

REMARKS

Entry of the foregoing, reexamination and further and favorable reconsideration of the subject application in light of the following remarks, pursuant to and consistent with 37 C.F.R. § 1.116, are respectfully requested.

Status

As is correctly reflected in the Office Action Summary, Claims 1-3, 9, 12, and 15-20 are pending. *See Final Official Action mailed February 8, 2005, Office Action Summary.* Claims 1 and 15-17 stand rejected. *Id.* Claims 2, 3, 9, 12, and 18-20 are objected to. *Id.*

As set forth in the Detailed Action, the Amendment and Reply filed November 5, 2004, has been entered. *See Final Official Action mailed February 8, 2005, Page 2, ¶ 1.* The former rejection of Claims 1-3, 9, 12, and 17-20 under 35 U.S.C. § 112, First Paragraph, has been withdrawn. *See Final Official Action mailed February 8, 2005, Page 2, ¶ 3.*

Summary of Claim Amendments

By the foregoing amendments, Applicants have amended Claims 1 and 15.

Specifically, Claim 1 has been amended to change the word "depend" to "attach," as recommended by the Examiner. *See Final Official Action mailed February 8, 2005, Page 2, ¶ 2.* Claim 1 has also been amended to add semicolons between the descriptions of the substituents of formula (I). These amendments are clerical in nature. Accordingly, no new matter has been added.

Claim 15 has been amended to delete the phrases "for repairing or combating ageing of the skin, whether this is light induced or chronological ageing, or for reducing actinic keratoses and pigmentations, or any pathologies associated with chronological or actinic ageing" and "for the treatment of dermatological complaints having an immunological component." Accordingly, no new matter has been added.

The amendments to Claim 1 and 15 are believed proper under 37 C.F.R. § 1.116 because they expressly comply with requests made in the Final Official Action and/or put the claims in better condition for appeal, if necessary.

Rejection Under 35 U.S.C. § 112, Second Paragraph

Claim 1 has been rejected under 35 U.S.C. § 112, Second Paragraph, as purportedly indefinite due to the term "depend." *See Final Official Action mailed February 8, 2005, Page 2, ¶ 2.* This rejection is respectfully traversed.

Not to acquiesce in the Examiner's rejection, but solely to expedite prosecution, by the foregoing amendments, Applicants have deleted from Claim 1 the term "depend" and have replaced it with the term "attach," as recommended by the Examiner. Applicants believe this amendment has rendered moot the foregoing rejection, and respectfully request withdrawal thereof.

Rejection Under 35 U.S.C. § 112, First Paragraph – Written Description and Enablement

Claims 15-17 were rejected under 35 U.S.C. § 112, First Paragraph, as purportedly lacking sufficient description and enablement. *See Final Official Action mailed February 8, 2005, Pages 2-3, ¶ 4.* These rejections are respectfully traversed.

Claim 15 Is NOT A “Reach-Through” Claim

Included with the Final Official Action in this case was a publication titled, “Reach Through Claims: Bust or Boom?” by Steve P. Lendaris and Robin M. Silva from Volume 4, No. 5, of the Intellectual Property Update provided by an intellectual property firm. The Examiner relies on this publication to characterize Claims 15-17 as “reach-through” claims. Applicants respectfully disagree.

The Lendaris publication states “[i]n general, ‘reach through’ claims attempt to capture the value of a discovery before it may be a full invention. For example, a research may identify a mechanism of disease action, such as a target protein involved in disease progression or the interaction of two proteins in the disease pathway. While these discoveries may allow the development of screening assays *to identify drug candidates, the actual products, the drugs themselves, have not yet been developed.* ‘Reach through’ claims are designed to cover these drugs or the use of them, ***prior to identification of the drugs themselves.***” (emphasis added).

Contrary to the reach-through claims described by Landaris and his colleague, Applicants ***have already identified the drugs*** used in Claims 15-17, *i.e.*, “the compound according to claim 1.” Because Applicants’ invention, as claimed in Claims 15-17, is concrete and not speculative, as were the purported inventions in

the highlighted *Rochester* and *Lilly* cases, Applicants submit that Claims 15-17 are not reach-through claims.

The Legal Tests of Written Description and Enablement

The written description requirement of 35 U.S.C. § 112, First Paragraph, asks whether the description "reasonably conveys to the artisan that the inventor had possession at the time of the later claimed subject matter." See *M.P.E.P.* § 2163.02 (citing *Ralston Purina Co. v. Far-Mar-Co., Inc.*, 772 F.2d 1570, 1575 (Fed. Cir. 1985)). The test of enablement is not whether any experimentation is necessary, but whether, if experimentation is necessary, it is undue. *M.P.E.P.* § 2164.01 (citing *In re Angstadt*, 537 F.2d 498, 504 (C.C.P.A. 1976)). Moreover, the fact that experimentation may be complex does not necessary make it undue, if the art typically engages in such experimentation. *M.P.E.P.* § 2164.01 (citing *In re Certain Limited-Charge Cell Culture Microcarriers*, 221 U.S.P.Q. 1165, 1174 (Int'l Trade Comm'n 1983)). Applicants respectfully submit that one of skill in the art would readily appreciate from the Specification that Applicants had possession of the subject matter of Claims 15-17 when the application was filed. In addition, Applicants respectfully submit that no *undue* experimentation is needed to arrive at the methods and compositions of Claims 15-17.

Applicants' Invention of Claims 15-17

Turning now to the rejections, not to acquiesce in the Examiner's rejection, but solely to expedite prosecution, by the foregoing amendments, Applicants have amended Claim 15 so as to delete the phrases "for repairing or combating ageing of the skin, whether this is light induced or chronological ageing, or for reducing actinic keratoses and pigmentations, or any pathologies associated with chronological or

actinic ageing" and "for the treatment of dermatological complaints having an immunological component." Applicants believe these amendments have rendered moot many of the concerns by the Examiner regarding the written description and enablement of Claims 15-17.¹

Regarding the other conditions treated in Claim 15, Applicants hereby provide for the Examiner's consideration three publications: (1) a table summarizing retinoid X receptor ("RXR") drugs, either marketed or in development; (2) a highlighted extract from the Annual Reports in Medicinal Chemistry, Volume 36; and (3) a highlighted copy of V. R. Atigadda *et al.*, "Conformationally Defined Retinoic Acid Analogues. 5. Large-Scale Synthesis and Mammary Cancer Chemopreventative Activity for (2*E*,4*E*,6*Z*,8*E*)-8-(3',4'-Dihydro-1'(2'*H*)-naphthalen-1'-ylidene)-3,7-dimethyl-2,4,6-octatrienoic Acid (9cUAB30)," 46 JOURNAL OF MEDICINAL CHEMISTRY 3766-3769 (2003).

As indicated in the Specification, some of the compounds are RXR agonists, whereas others are RXR antagonists. As indicated by the first publication, there are several RXR drugs used in the treatment of different cancers, psoriasis, and eczema. As indicated by the second publication, the RXR agonist Bexarotene is used in treating T-cell lymphoma (see Page 298). The third publication indicates that RXR agonists may be efficient in mammary cancer chemoprevention. This publication indicates that the RXR agonist 9cUAB30 of formula (I) "is very effective in the prevention of *N*-methyl-*N*-nitrosourea induced mammary cancers in rats without signs of toxicity." Taken together, these publications demonstrate the efficiency of RXR retinoids in the treatment of cancers, psoriasis, and eczema.

¹ Applicants wish to draw the Examiner's attention to the fact that Claims 16 and 17 depend on Claim 1, not Claim 15.

In view of the foregoing and the totality of information provided in Applicants' disclosure, Applicants submit that those of skill in the art would readily appreciate that Applicants were in possession of the invention of Claims 15-17 and that those skilled in the art can readily practice the invention of Claims 15-17. Accordingly, Applicants respectfully request withdrawal of the 35 U.S.C. § 112, First Paragraph, written description and enablement rejections of Claims 15-17.

CONCLUSION

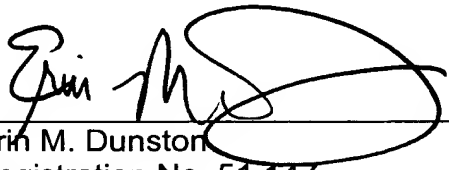
From the foregoing, further and favorable action in the form of a Notice of Allowance is respectfully requested and such action is earnestly solicited.

In the event that there are any questions relating to this Amendment and Reply Following Final Rejection, or the application in general, it would be greatly appreciated if the Examiner would telephone the undersigned attorney concerning such questions so that the prosecution of this application may be expedited.

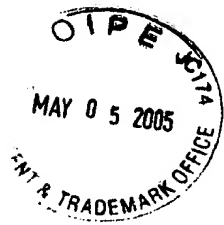
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Date: May 5, 2005

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RXR+ Retinoids for Treatment of Cancer

1- Marketed Drugs

Drug Names	Company	Indications
Panretin Gel 0.1% (alitretinoin; 9-cis retinoic acid)	Ligand	cutaneous lesions of AIDS-related Kaposi's sarcoma
Targretin Gel 1 % (bexarotene)	Ligand	cutaneous T cell lymphoma (CTCL)
Targretin Capsules (bexarotene)	Ligand	refractory T cell lymphoma (CTCL)

2- Drugs in Development

Drug Names	Company	Indications
Panretin Oral Capsules (alitretinoin)	Ligand	bronchial metaplasia
Targretin Oral Capsules (bexarotene)	Ligand	psoriasis
Targretin Gel	Ligand	psoriasis
MC-1035 (structure not known)	Maxocore Pharmaceuticals	lung cancer

Note: Topical alitretinoin (Basilea) and oral bexarotene (Ligand) are in phase II / III development for treatment of severe, chronic contact eczema of the hands.

Conformationally Defined Retinoic Acid Analogues. 5. Large-Scale Synthesis and Mammary Cancer Chemopreventive Activity for (2*E*,4*E*,6*Z*,8*E*)-3-(3',4'-Dihydro-1'(2'*H*)-naphthalen-1'-ylidene)-3,7-dimethyl-2,4,6-octatrienoic Acid (9cUAB30)

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Retinoids that activate the nuclear retinoid X receptors (RXRs) display potential for chemoprevention of breast cancer. We previously reported that 9cUAB30 (**1**) is an RXR-selective retinoid. To explore its *in vivo* chemopreventive activity, multigram quantities of **1** were needed. Here, we describe a modified synthesis that yields up to 100 g of **1**. We further demonstrate that **1** is very effective in the prevention of *N*-methyl-*N*-nitrosourea induced mammary cancers in rats without signs of toxicity.

In humans, the estrogen receptor antagonist Tamoxifen is used as adjuvant therapy for high-risk patients,¹ and currently it is the only chemopreventive agent for breast cancer approved by the Food and Drug Administration. Even though the benefits of Tamoxifen are substantial, long-term administration is not without risks. Women who undergo Tamoxifen therapy have higher levels of endometrial cancers, since Tamoxifen acts as an agonist rather than an antagonist in the endometrium.² Other chemopreventive agents that can be used alone or in combination with Tamoxifen are needed. Since retinoids are capable of inducing apoptosis and cell differentiation, these agents have been explored for their chemopreventive effects. The efficacy of retinoids as chemopreventive agents has been demonstrated for numerous animal models of carcinogenesis including skin, breast, oral cavity, lung, hepatic, gastrointestinal, prostatic, and urinary bladder cancers.³

Anzano et al.⁴ showed that 9-*cis*-retinoic acid is a potent retinoid for the prevention of *N*-methyl-*N*-nitrosourea (MNU) induced mammary carcinogenesis. However, since the effective dose is near the toxic dose, this agent may not be suitable for long-term administration needed in chemoprevention. The all-*trans*-retinoic acid is much less effective in these assays, suggesting that retinoids that activate the retinoid X receptors (RXRs) may be more efficacious in mammary cancer chemoprevention. Gottardis et al.⁵ demonstrated that Targretin, an RXR-selective ligand, prevented the appearance of MNU-induced rat mammary tumors. Furthermore, this retinoid had less toxicity than 9-*cis*-retinoic acid. When either 9-*cis*-retinoic acid or Targretin was used in combination with Tamoxifen, increased chemopreventive efficacy was observed over either agent alone.⁶ Recently we reported a new RXR-selective ret-

inoid, **1**,⁷ which is a conformationally constrained analogue of 9-*cis*-retinoic acid that locks the tetraene chain in a defined conformation. The retinoid **1** binds to RXRs and efficiently induces transcription mediated by these receptors over retinoic acid receptors (RARs). To evaluate the effectiveness of this retinoid in reducing the MNU-initiated mammary cancers in rats, multigram quantities of **1** were required. Here, we report a new synthesis of **1** suitable for the efficient preparation of multigram quantities and the results of mammary cancer chemopreventive assays and toxicity in rats.

Chemistry

Our previously reported⁷ synthesis of **1** is shown in Scheme 1. In this procedure, a Reformatsky reaction between α -tetralone (**2**) and ethyl 4-bromo-3-methyl-2-butenolate (**3**) directly provided the acid **5** (86%). Reduction of the acid to the alcohol **6** (67%) [1:1 (9*Z*) to (all-*E*) mixture], followed by oxidation, provided the aldehyde (9*Z*)-**7** (69%) [1:1 (9*Z*) to (all-*E*) mixture; pure (9*Z*) isomer obtained by flash chromatography]. A Horner–Emmons condensation between aldehyde (9*Z*)-**7** and triethyl phosphonosenecioate (**8**) provided the ester **9** (78%) as a 2:1 mixture of (9*Z*) and (9*Z*,13*Z*) isomers. These isomers were separated by HPLC, and the desired (9*Z*)-**9** was hydrolyzed under basic conditions (98% yield) to give **1**.

While satisfactory for a small scale, this methodology was not amenable for large-scale synthesis of multigram quantities needed for the chemoprevention studies. The oxidation of the alcohol to the aldehyde required the use of large amounts of MnO₂ and molecular sieves, and the purification became extremely tedious at larger scale. Oxidation of 100 g of alcohol **6** would require 1 kg of MnO₂ and 0.5 kg of powdered molecular sieves. Separation of the product would require washing the MnO₂ with about 15 L of solvent, and during this process, a considerable amount of aldehyde decomposes. At this scale, the yield of the aldehyde is expected to be considerably lower than 69%. Furthermore, the Horner–Emmons reaction resulted in a 2:1 mixture of

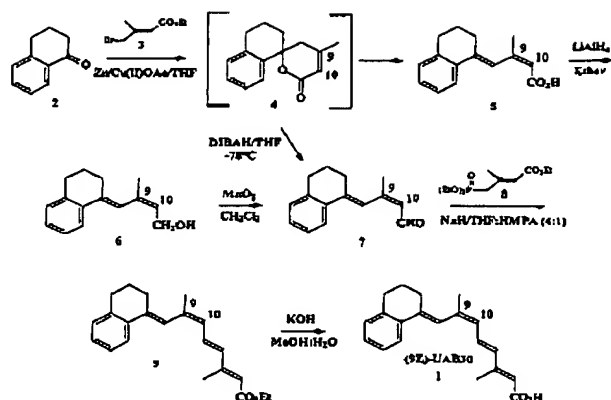
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Brief Articles

Scheme 1



(9Z)-9 and (9Z,13Z)-9. This is another problematic step because the separation of the two isomers requires HPLC, which is impractical for large scales, and because one-third of the aldehyde 7 is wasted by conversion to the undesired di-Z ester 9. These limitations prompted us to develop an alternative synthetic methodology more amenable for a large-scale synthesis.

The modified synthetic methodology is also depicted in Scheme 1. As shown, the first step also involved a Reformatsky reaction between 1-tetralone (2) and ethyl 4-bromo-3-methyl-2-butenate (3) in the presence of Zn and Cu(OAc)₂ in THF.⁸ However, conditions were used to favor the formation of the intermediate δ -lactone 4. This reaction was performed on a 100 g scale to yield approximately 100 g (70%) of 4. Another major change was the controlled reduction of 4 by DIBALH, followed by ring opening and elimination, to provide the aldehyde 7 (75%) [5:1 (9Z) to (all-E)]. The isomers were readily separated on flash silica, and triethyl phosphonosenecioate 8 was used to olefinate (9Z)-7 under modified Horner–Emmons conditions to produce the ester 9. Under these conditions, the use of excess HMPA as the solvent resulted in the desired ester 9 as a 9:1 mixture of (9Z,13E)-9 and (9Z,13Z)-9, which were separated by selective crystallization. Pure (9Z,13E)-9 was hydrolyzed under basic conditions to give the pure acid 1 in 78% yield.

Biology

Female Sprague–Dawley rats were obtained from Harlan Sprague–Dawley, Inc., at 23 days of age. The animals were housed five per cage and maintained on a Teklad (4%) rodent diet throughout the study. At 50 days of age, the rats received one intravenous injection of MNU via the jugular vein. MNU was purchased from the NCI Chemical Repository. The animals ($n = 20$ per group) were administered with 1 or 9-*cis*-retinoic acid

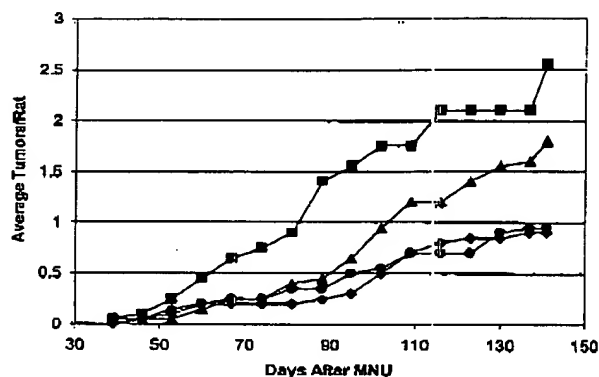


Figure 1. Average tumor formation versus days after MNU administration for female rats fed a 4% Teklad diet with 9cUAB30 (200 mg/kg diet, diamond; 100 mg/kg diet, triangle), with 9-*cis*-retinoic acid (100 mg/kg diet, circle), or without retinoid (MNU control, square).

in the diet continuously after MNU injection, beginning at 53 days of age (3 days after the carcinogen). The compounds were incorporated into the diet using a Patterson–Kelly liquid–solid blender. Stability of the retinoids during feeding was verified by HPLC. The groups were as follows: group 1, 1 (200 mg/kg diet); group 2, 1 (100 mg/kg diet); group 3, 9-*cis*-retinoic acid (100 mg/kg diet); group 4, diet only. The rats were weighed once per week, palpated for mammary tumors twice per week, and checked daily for signs of toxicity. The study was terminated 140 days after MNU treatment. All mammary tumors were histologically classified as adenocarcinomas.

Results and Discussion

1 and 9-*cis*-retinoic acid were tested in the MNU-induced mammary cancer model currently used by the National Cancer Institute to evaluate the efficacy of chemopreventive agents.⁴ At termination of the study, the average number of mammary cancers per rat for the control group was 2.6. 1 at the 200 and 100 mg/kg diet dose levels reduced the number of mammary cancers by 63% and 29%, respectively (Figure 1). The use of 9-*cis*-retinoic acid at the 100 mg/kg diet dose level (a level just below the toxic dose for this compound) decreased the multiplicity of cancers by 65%. Both the high dose of 1 and 9-*cis*-retinoic acid greatly delayed the time of appearance of the mammary cancers (Figure 1). As shown in Table 1, retinoid 1 did not alter body or liver weights of the rats significantly at either dose level. Other signs of retinoid clinical toxicity were not observed. Since mammary cancers are highly sensitive to ovarian hormone changes, we measured the weight of the ovaries and uterus at the end of the study and

Table 1

retinoid	final body weight (g)	organ weight (g) per 100 g of body weight		
		liver	uterus	ovaries
MNU control	269 ± 9 ^a	3.22 ± 0.05	0.18 ± 0.03	0.046 ± 0.002
1 (100 mg/kg diet)	271 ± 3	2.96 ± 0.07	0.16 ± 0.01	0.046 ± 0.002
1 (200 mg/kg diet)	272 ± 9	3.11 ± 0.09	0.21 ± 0.02	0.052 ± 0.005
9- <i>cis</i> -retinoic acid (100 mg/kg diet)	254 ± 7	3.48 ± 0.09	0.18 ± 0.01	0.040 ± 0.003

^a Values are the mean ± SEM; $N = 5$. Statistical differences between groups were not observed.

monitored the estrus cycles of the rats in the various groups. Since major differences between groups were not observed, it appeared that 1 modified the carcinogenic process by a mechanism other than through hormonal modifications. Anzano et al. reported similar efficacy for 9-*cis*-retinoic acid in mammary cancer chemoprevention using this MNU assay.⁴

Conclusions

Synthetic methodology suitable for generating 1, and presumably related retinoids, on a multigram scale was developed. Retinoid 1 was found to be highly active in the prevention of mammary carcinogenesis in rats and was completely nontoxic at the highest tested dose when administered orally. This and related retinoids warrant further evaluation as potential therapeutic agents in mammary chemoprevention.

Experimental Section

Melting points were obtained on an electrothermal melting point apparatus and are uncorrected. ¹H and ¹³C NMR spectra were recorded on a Bruker ARX 300 spectrometer. UV/vis spectra were recorded on a Varian Cary 100 Conc spectrophotometer in methanol. IR spectra were recorded using a Bomem MB series FT IR spectrometer. Mass spectra were recorded on a MicroMass platform LCZ spectrometer. Atlantic Microlabs of Atlanta, GA, provided combustion analyses. Solvents and liquid starting materials were distilled prior to use. Reactions and purifications were conducted with deoxygenated solvents under inert gas (N₂) and in subdued lighting. Flash chromatography was performed using Selecto Scientific silica gel (40 μm). Ethyl 4-bromo-3-methylbut-2-enoate (3) was prepared by the reaction of ethyl 3,3-dimethylacrylate with *N*-bromosuccinimide.^{9–11} Triethyl phosphonoselenoate (8) was prepared via the Arbuzov reaction.¹² Tetrahydrofuran was distilled from sodium metal/benzophenone ketyl. Diethyl ether, benzene, and dichloromethane were purchased from Fischer as anhydrous solvents. HMPA was distilled from calcium hydride.

7,8-Benzo-4-methyl-1-oxaspiro[5.5]undec-3-en-2-one (4). A mixture of zinc dust (150 g) (<10 μm, Aldrich, catalog no. 20,998-8) and copper(II) acetate monohydrate (15 g, Acros) in 500 mL of glacial acetic acid was stirred under nitrogen for 1 h in a 1000 mL one-neck, round-bottomed flask. The mixture was diluted with anhydrous ether (500 mL) and filtered with suction, and the Zn–Cu complex was washed successively with anhydrous ether (3 × 300 mL) and dry benzene (3 × 300 mL). The mixture was then transferred into a flame-dried 2000 mL three-neck flask fitted with a nitrogen inlet, condenser, and addition funnel. Freshly distilled THF (distilled from Na/benzophenone) (200 mL) was added to the flask, which was heated to about 90 °C in an oil bath. The reaction mixture was then treated dropwise with a solution of tetralone 2 (100.0 g, 684.9 mmol, freshly distilled) and bromoester 3 (220.0 g, 1063 mmol, freshly distilled) in 100 mL of THF (dry). Vigorous bubbling occurred during the addition. The mixture was stirred at reflux for an additional 3.5 h. The reaction mixture was cooled to room temperature, and water (200 mL) and HCl (2 N, 500 mL) were added. The mixture was diluted with 1000 mL of ether and filtered, and the acid layer was separated. The organic layer was washed with water (2 × 200 mL), NaOH (1 N, 2 × 250 mL), and brine (2 × 250 mL). It was then dried (Na₂SO₄) and evaporated to give an oil. This oil was subjected to distillation on a high-vacuum pump (0.1 mm) at 60 °C. The distillate was discarded, and the remaining thick oily residue solidified upon addition of hexanes. This mixture was cooled, filtered, and washed with hexanes to give 108 g (69.2%) of 4 (*R*_f = 0.3, 50:50 ether/hexane) as a white solid: mp 67–69 °C; MS *m/z* 229 (*M*⁺); ¹H NMR (CDCl₃) δ 7.5–7.54 (m, 1H), 7.2–7.25 (m, 2H), 7.07–7.1 (m, 1H), 5.92 (s, 1H), 2.7–2.9 (m, 3H), 2.5 (d, 1H), 1.98–2.23 (m, 3H), 2.01 (s, 3H), 1.67–1.78 (m, 1H).

(2Z,4E)-4-(3',4'-Dihydro-1'(2'H)-naphthalen-1'ylidene)-3-methyl-2-butenal (7). To a flame-dried three-neck, round-bottomed flask fitted with a nitrogen inlet, addition funnel, and rubber septum was added lactone 4 (20.0 g, 87.6 mmol). To this was added 400 mL of THF (freshly distilled from Na/benzophenone). The resulting solution was cooled to –78 °C in a dry ice/acetone bath. The reaction mixture was treated with diisobutylaluminum hydride (88.0 mL, 87.6 mmol, 1 N solution in THF, Aldrich) dropwise over a period of 45 min. After 2 h of stirring at –78 °C, an additional amount of DIBALH (8.80 mL, 8.76 mmol) was added dropwise. After an additional 3 h of stirring, more DIBALH (8.80 mL, 8.76 mmol) was added dropwise, and stirring continued at –78 °C for an additional 20 h. The reaction mixture was quenched with 20 mL of water, and the dry ice bath was removed. After reaching room temperature, the mixture was warmed to about 35 °C in a water bath and 60 mL of 18% HCl was added. The mixture was stirred for 10 min at 35–40 °C. The reaction mixture was diluted with ether (200 mL), washed with water (3 × 100 mL) and brine (2 × 100 mL), dried (Na₂SO₄), and evaporated (rotary evaporator, water bath temperature of <35 °C) to give 18 g of an oil, which was purified by column chromatography (silica gel, 40 cm × 7 cm, 1:6 ether/hexanes, all column solvents purged with nitrogen) to give 10 g of (9Z)-7 (*R*_f = 0.3) and 2.5 g of (all-*E*)-7 (*R*_f = 0.25) (75% combined yield). The (9Z)-7 was crystallized from hexanes/ether: mp 65–66 °C; IR 1662 (C=O), 1609 (C=C) cm⁻¹; UV λ_{max} 295 (ε 6000); MS *m/z* 213 (*M*⁺ + H); ¹H NMR (CDCl₃) δ 9.64 (d, 1H), 7.64 (m, 1H), 7.13–7.25 (m, 3H), 6.57 (s, 1H), 6.0 (d, 1H), 2.36 (t, 2H), 2.50 (t, 2H), 2.09 (s, 3H), 1.82–1.90 (m, 2H).

(2E,4E,6Z,8E)-Ethyl 8-(3',4'-Dihydro-1'(2'H)-naphthalen-1'ylidene)-3,7-dimethyl-2,4,6-octatrienoate (9). Sodium hydride (60% suspension in mineral oil, 2.95 g, 73.8 mmol) was placed in a flame-dried three-neck, round-bottomed flask fitted with a nitrogen inlet, addition funnel, and rubber septum. Freshly distilled THF (from Na/benzophenone, 400 mL) was added, followed by freshly distilled 8 (19.45 g, 73.67 mmol). The resulting brown mixture was stirred for 15 min, and freshly distilled HMPA (50 mL) was introduced through a syringe. The flask was covered with aluminum foil, and stirring was continued for 15 min. The aldehyde 7 (14.20 g, 66.98 mmol) in 100 mL of dry THF was added dropwise from the addition funnel (covered with aluminum foil). The reaction mixture was stirred for an additional 2.5 h, was quenched with 50 mL of water, and then diluted with 500 mL of ether. The aqueous layer was separated and washed with 100 mL of ether. The combined organic layers were washed with brine (2 × 150 mL), dried (Na₂SO₄), and evaporated to give a crude oil (35 g), which was suspended in methanol (75 mL, degassed with nitrogen). Ether was added until the mixture was homogeneous (about 20 mL), and the solution was cooled overnight at 0 °C to give a crystalline solid. This solid was filtered, washed with methanol, and dried to give 14 g of pure product (9Z)-9 as one isomer: mp 64–65 °C; IR 1706 (C=O), 1602 (C=C) cm⁻¹; UV λ_{max} 328 nm (ε 29 300); MS *m/z* 323 (*M*⁺ + H); ¹H NMR (CDCl₃) δ 7.62–7.68 (m, 1H), 7.11–7.22 (m, 3H), 6.65 (dd, 1H), 6.5 (s, 1H), 6.23 (d, 1H), 6.1 (d, 1H), 5.75 (s, 1H), 4.15 (q, 2H), 2.85 (t, 2H), 2.40 (dt, 2H), 2.22 (s, 3H), 1.97 (s, 3H), 1.78–1.87 (m, 2H), 1.27 (t, 3H).

(2E,4E,6Z,8E)-8-(3',4'-Dihydro-1'(2'H)-naphthalen-1'ylidene)-3,7-dimethyl-2,4,6-octatrienoic Acid (9cUAB30, 1). Ester 9 (12.00 g, 37.26 mmol) was suspended in methanol (640 mL, degassed with nitrogen) and warmed to about 60 °C. This mixture was treated with KOH solution (20.90 g, 372.7 mmol, in 220 mL of distilled and degassed water). The resulting mixture was stirred at reflux for 1 h, cooled to 0 °C in an ice bath, and diluted with 300 mL of ice-cold water. The mixture was slowly acidified with ice-cold 2 N HCl to about pH 2. The resulting precipitate was filtered, and the solid was redissolved in 500 mL of ether. The organic solution was washed with brine (3 × 150 mL), dried (Na₂SO₄), and concentrated on a rotary evaporator to about 75 mL of volume. The residual solution was diluted with 100 mL of degassed hexanes and cooled at 0 °C for about 12 h. The resulting yellow crystals

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were filtered and dried to give 8.5 g (78%) of pure (9Z)-1 (9Z-9cUAB30): mp 175–176 °C; IR 1672 (C=O), 1594 (C=C) cm^{-1} ; UV λ_{max} 328 nm (ϵ 30 200); MS m/z 295 ($\text{M}^+ + \text{H}$); ^1H NMR (CDCl_3) δ 11.00 (br, 1H), 7.6–7.67 (m, 1H), 7.15–7.21 (m, 2H), 7.11–7.14 (m, 1H), 6.68 (dd, 1H), 6.47 (s, 1H), 6.25 (d, 1H), 6.12 (d, 1H), 5.77 (s, 1H), 2.85 (t, 2H), 2.40 (dt, 2H), 2.22 (s, 3H), 1.98 (s, 3H), 1.79–1.87 (m, 2H).

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Section VII—Trends and Perspectives

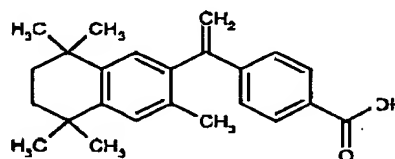
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and inhibit LTD₄ in tracheal smooth muscle and ileum, IL-5 production by human peripheral blood mononuclear cells as well as eosinophil infiltration in the airway and peripheral blood. As a consequence, it is currently being developed against other allergic and respiratory disorders.



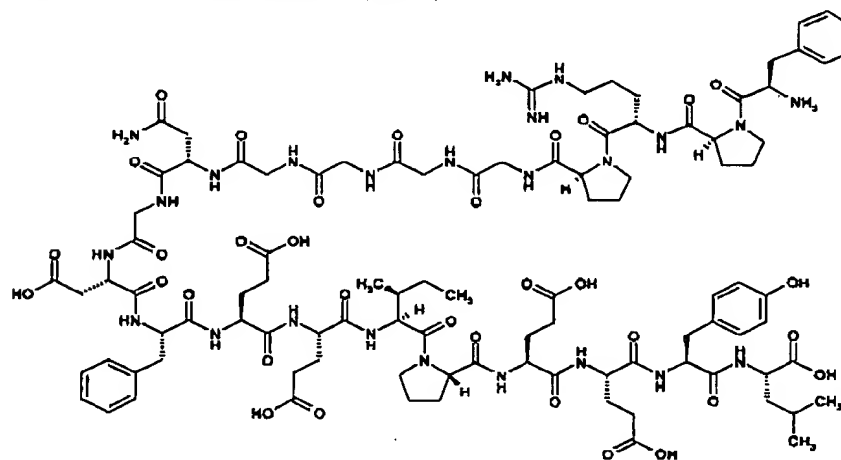
Bexarotene (anticancer) (26-29)

Country of Origin : US
 Originator : Ligand
 First Introduction : US
 Introduced by : Ligand
 Trade Name : Targretin
 CAS Registry No : 153559-49-0
 Molecular Weight : 348.49



Bexarotene was launched in the US for the treatment of manifestations of cutaneous T-cell lymphoma in patients who are refractory to at least one prior systemic therapy. The four step synthesis of bexarotene involves a double Friedel-Craft alkylation of toluene with 2,5-dichloro-2,5-dimethylhexane followed by acylation with monomethylterephthalic acid chloride, then Wittig methylenation. Bexarotene is the first retinoid X receptor (RXR) agonist to be selective versus retinoid A receptors (RAR). Its activation of the three RXR α , β , γ isoforms induces cell differentiation and apoptosis and inhibits cell proliferation in several models of cancer. In phase II/III clinical trials, 48% of patients with refractory or persistent early-stage cutaneous T-cell lymphoma achieved a complete or partial response when treated with 300 mg/m²/day of bexarotene. It was shown in phase I trials that this second-generation retinoid was substantially less toxic than the broad-spectrum or RAR-selective retinoids.

Bivalirudin (antithrombotic) (30-37)



Country of Origin : US
 Originator : Biogen
 First Introduction : New Zealand
 Introduced by : The Medicines Company

Trade Name : Angiomax
 CAS Registry No : 128270-60-0
 Molecular Weight : 2179.3